Pathogenesis and Treatment of Human Immunodeficiency Virus and Chronic Hepatitis C Virus Infection

THE HEPATITIS C VIRUS (HCV) IS A SINGLE-STRANDED, ENVELOPED VIRUS OF THE FLAVIVIRIDAE FAMILY, IDENTIFIED IN 1989. USUALLY, HCV PERSISTs AS A CHRONIC INFECTION, POTENTIALLY LEADING TO CIRRHOSIS, LIVER FAILURE, AND HEPATOCELLULAR CARCINOMA (HCC) IF UNTREATED. IN THE UNITED STATES AND EUROPE, LIVER DISEASE SECONDARY TO HCV IS THE LEADING INDICATION FOR LIVER TRANSPLANTATION.

Due to overlapping transmission routes, HCV coinfection is common among HIV-infected persons. In the United States and Europe, HCV-associated end-stage liver disease has become a leading cause of death among HIV-infected persons (Bica, 2001; Rosenthal, 2003).

Virology

THE RNA GENOME IS APPROXIMATELY 9400 NUCLEOTIDES IN LENGTH, comprising 1 long open reading frame that encodes a polyprotein of 3010-3033 amino acids. This protein is cleaved into functionally distinct polypeptides during or after translation. The nucleocapsid and envelope proteins are encoded at the 5' end of the genome, while the nonstructural elements are located downstream. There are hypervariable regions (particularly in the E1 and E2 domains that code for envelope glycoproteins) that may be important antigenic sites. Their variability may be central in persistence of infection and immunopathogenesis. The hepatitis C virus replicates via an error-prone RNA-dependent RNA polymerase. Daily production of HCV exceeds 12 billion virions. Infections with HCV typically occur as a mixture of closely related viral populations known as quasispecies. There is a high degree of genetic variability, and the virus has evolved to optimize fitness in different hosts. There are at least 6 HCV genotypes, and hundreds of subtypes. Genotypes 1, 2, and 3 are the most common in the United States and Europe.

The HCV genotype is the most important pretherapy virologic predictor of outcome, regardless of HIV status (Fried, 2002; Manns, 2001; Carrat, 2004; Chung, 2004; Hadziyannis, 2004; Laguno, 2004; Torriani, 2004). Treatment is less effective for persons infected with HCV genotype 1, which is predominant in the United States (Blatt, 2000).

Epidemiology

CHRONIC HCV INFECTION IS AN IMPORTANT PUBLIC HEALTH CONCERN. Worldwide, 170 to 400 million people are thought to be infected with HCV, and a higher prevalence is found in Asia and Africa (Tossing, 2005). Population prevalence data are unavailable for many countries, but it is believed that 2% to 3% of the world's population is chronically infected. HCV is one of the 10 leading causes of infectious disease deaths worldwide, annually accounting for 250,000 deaths (Perz, 2006).

Globally, an estimated 4 to 5 million people are coinfected with HIV/HCV (Alter, 2006). In the United States, 20%–30% (Sulkowski, 2000) and in Europe, 34% (Rockstroh, 2004) of all HCV-infected persons are coinfected with hepatitis C, with much higher rates in hemophiliacs and injection drug users (IDUs). Among cohorts of HIV-seropositive IDUs, prevalence ranges from 70% to 90% (Garfein, 1996; Sherman, 2002; Bowker, 2004; Miller, 2004; Mohsen, 2005). Even though the rate of sexual transmission of HCV in heterosexual monogamous couples is exceedingly low, sexual transmission in both HIV-infected and -uninfected men who have sex with men (MSM) has now been documented (Danta, 2007).

In developed countries, individuals at risk of HCV infection include:

- Current and former injection drug users
- Health care workers who have had an occupational exposure to blood or blood products
- Individuals on hemodialysis, if facility does not practice adequate infection control
- Those who engage in high-risk sexual practices. (see Mark Danta, page 2 in this issue)
- Blood transfusion prior to 1992
- Sharing of paraphernalia in intranasal cocaine users
- Maternal/child transmission (C-section is recommended for HIV/HCV coinfected women)
- Tattooing, when under unsterile conditions such as in prison with shared equipment, ink, inkwells

In developing countries, sources of HCV infection include all of the above and:

- Transfusions of unscreened blood
- Those who receive unsafe injections, or other parenteral exposure to blood
- Use of blood-contaminated implements for circumcision or surgery
- Traditional scarification
- Acupuncture under unsterile conditions
- Ear piercing
Natural History and Pathogenesis

Most cases of chronic HCV are asymptomatic (Sanantonio, 2003) and not preceded by an episode of clinically apparent jaundice (Calleri, 2007). In 15% to 45% of exposed individuals, acute HCV disease completely resolves, with clearance of HCV RNA from serum within 4 months (Di Bisceglie, 1991; Tremolada, 1992). Hepatitis C viral RNA becomes detectable in serum 7 to 21 days after exposure, while HCV antibodies are present from 20 to 150 days. Less than 20% of those infected have jaundice, and this usually correlates with RNA titers. In addition, a clinical syndrome may occur, consisting of jaundice, nausea, fatigue, myalgias, low-grade fevers, and upper abdominal discomfort. When present, it occurs within 2 to 12 weeks after exposure, lasting up to 12 weeks, and may not be related to viral titers (Orland, 2001). HCV-specific CD4+ and CD8+ T cells appear within 2 to 3 months following acute infection. A strong T cell response, with increased production of interferon-gamma (INF-γ) and interleukin 2 (IL-2), was characteristic in HCV-monoinfected subjects who cleared viremia.

Approximately 55% to 85% of exposed individuals fail to spontaneously clear the virus and are at risk for liver damage as well as extrapathologic manifestations of HCV. Progression of HCV-induced liver disease has been associated with many systemic diseases and their complications, including autoimmune hepatitis, cryoglobulinemia, porphyria cutanea tarda, lymphocytic sialoadenitis and membranous glomerulonephritis. Data in support of a link between non-Hodgkins lymphoma and HCV (Gisbert, 2004) also exist but may be limited to MSM (Franceschi, 2006).

The pathogenetic mechanisms that result in chronic hepatitis are unknown; Tsai and colleagues proposed that abnormalities in early events, involving innate immunity, may lead to the impaired cellular immunity responses seen in those who develop persistent infection (Tsai, 1997). Thus, loss of sustained CD4+ reaction (Gerlach, 1999) and lower IFN-γ response to NS 3-5 proteins (Danta, 2006), lead to chronicity. In HIV-HCV coinfection, there is increased apoptosis of CD4+ and CD8+ naïve and memory cells, suggesting a permissive effect of HIV, regardless of viral load or CD4 counts, towards establishing chronic HCV infection. Furthermore, reduced activation of CD8+ memory cells is noted in coinfected subjects (Yonkers, 2006).

In chronic HCV infection, fibrosis results from the activation of hepatic stellate cells by cytokines and other signalling molecules induced by the inflammatory process. These produce and deposit extracellular matrix proteins. Fibrosis begins around the portal tracts and gradually extends into the lobules towards the central veins. Hepatic steatosis is a concurrent factor in the progression to advanced fibrosis. Steatosis is associated with HCV genotype 3, and in patients with other genotypes, steatosis is associated with metabolic factors such as higher body mass index, type 2 diabetes, and hyperlipidemia (Castera, 2006). HCV, with or without HIV, increases the risk of insulin resistance and diabetes (Duong, 2001; Mehta, 2003). The use of a protease inhibitor is an additional independent risk factor for developing hyperglycemia (Mehta, 2003). Insulin resistance is a common denominator for steatosis, fibrosis, and elevated circulating tumor necrosis factor (TNF), which adversely affect sustained virologic response (SVR) rates of genotype 1 HCV-infected patients (Romero-Gomez, 2005; 2006). In contrast to monoinfected patients, insulin resistance and diabetes were not contributing factors to fibrosis progression in HCV/HIV coinfected patients (Merchant, 2006; Monto, 2006), although a recent report has shown hyperglycemic patients to be more likely to have advanced fibrosis (Barreiro, 2006).

At least 20% of those with chronic hepatitis will progress to cirrhosis after 20 years, increasing the risk of liver failure and HCC. The disease can more rapidly progress to cirrhosis in those with other hepatotoxic risk factors, such as alcohol consumption of more than 50 grams per day, aging, and HBV and/or HIV coinfection. Approximately 10% to 20% of patients become cirrhotic within 10 years of infection. The initial presentation of older patients without access to medical care may be with advanced liver disease, cirrhosis, and/or HCC. Currently, HCV-induced cirrhosis is the most common indication for liver transplantation in the United States and Europe.

Effects of Coinfection

Coinfection with HIV affects the natural history of HCV infection. Lower clearance rates (5% to 10%) are seen in HIV-seropositive individuals with acute HCV. HIV accelerates HCV disease progression. Cirrhosis is more prevalent among HIV-positive than -negative patients (Di Martino, 2001) and is emerging as a major cause of morbidity and mortality in patients with HIV/HCV coinfection (Rosenthal, 2003). Liver disease is now among the leading causes of hospital admissions for patients with HIV infection, and timely evaluation of HCV in these patients is crucial.

HCV is an independent risk factor for highly active antiretroviral therapy (HAART)-associated hepatotoxicity. This effect is generally outweighed by a reduction in the overall risk of liver-related mortality when HAART is administered; only 4% of HAART-treated patients require discontinuation of their HIV medication (Cooper, 2006). In a recent study, 2.7% of liver mortality was attributed to antiretrovirals, while HCV was responsible for 10% of liver-related events in the HAART era (Weber, 2006). Immune status and virologic control are critical in fibrogenesis. Factors that adversely impact fibrosis progression (Verma, 2006) include: decline of more than 10% in CD4 (Schiavini, 2006); nadir CD4 (Monto, 2006); current CD4 below 200 cells per mm³ (Di Martino, 2001), and HIV RNA above 50 copies per mL (Hernandez, 2006). Maintaining a CD4 count above 500 cells per mm³ may be associated with lower fibrosis progression rates. Successful HIV suppression in patients with CD4 levels below 500 cells per mm³ has also been associated with slower progression of fibrosis (Brau, 2006). Beneficial effects may be seen with any suppressive HAART regimen (Verma, 2006).

Antiretroviral agents for coinfected patients should be carefully selected, since some are more likely to induce hepatotoxicity. Liver enzyme levels should be routinely monitored, and therapy stopped if symptoms develop or if liver function tests rise to greater than 5 times normal or to above 3.5 times the baseline values. In addition, other etiologies such as ethanol abuse, acute cholecystitis, or infection with hepatotropic viruses, like Epstein-Barr or cytomegalovirus (CMV) have to be ruled out (Sulkowski, 2003)

Stavudine and didanosine, nevirapine, and tipranavir—the latter in combination with enfuvirtide—have been associated with serious liver damage in persons with HIV alone (Maida, 2006). Stavudine and didanosine may deplete hepatocellular mitochondrial RNA, and have been associated with hepatic steatosis in HIV/HCV coinfected patients (Mc Govern, 2006; Sulkowski, 2005; Walker, 2004). The effect of HCV on HIV progression is poorly understood and is beyond the scope of this review.

Diagnoses

Anti-HCV antibody is usually detectable within 3 weeks of exposure, whereas HCV-RNA is detectable in blood 1 to 3 weeks after exposure (Orland, 2001; Netski, 2005). Patients with positive serology for antibodies against HCV should have a qualitative RNA level measured to confirm viremia. Up to 5.5% of coinfected indi-
individuals will display seronegative chronic HCV infection, in which there is negative serology and detectable HCV RNA (Bonacini, 2001). Elevated ALT, history of intravenous drug use and CD4 levels below 200 cells per mm³ are predictors of seronegative HCV (Chamiie, 2006), and may indicate further screening for HCV RNA.

Two polymerase chain reaction (PCR)-based tests for qualitative detection of HCV RNA are currently approved by the FDA: Amplicor Hepatitis C Virus Test, version 2.0, and Cobas Amplicor Hepatitis C Virus Test, version 2.0 (Roche Molecular Systems, Branchburg, NJ), which have lower limits of detection of approximately 50 IU per mL. Of the quantitative tests (Table 1), only Versant HCV RNA version no. 3.0 is approved by the FDA. If HCV RNA is detected, baseline serum aminotransferases, bilirubin, alkaline phosphatase, prothrombin time, complete blood count (CBC) with differential, creatinine, and thyroid-stimulating hormone (TSH) should be measured in preparation for treatment. It is advisable to rule out other causes of liver disease such as hemochromatosis or autoimmune hepatitis.

Serum aminotransferases (AST and ALT) remain abnormal after 12 months in 60% to 85% of patients with posttransfusion or sporadic hepatitis. These enzymes decline from the peak values encountered in the acute phase of the disease, but typically remain abnormal by 2- to 8-fold. Serum ALT concentrations may fluctuate during the course of the disease, but they can also be intermittently or consistently normal. As chronic disease progresses, laboratory values continue to become more abnormal. Serum AST greater than ALT, hypoalbuminemia, thrombocytopenia, and prolonged prothrombin time all suggest cirrhosis.

A liver biopsy is helpful in grading the degree of inflammation and staging the degree of fibrosis. Biopsy has prognostic value, since all patients with initial periportal fibrosis are likely to develop cirrhosis after 2 decades of untreated infection (Yano, 1996). In patients with less-severe histologic disease who may never develop cirrhosis, careful clinical monitoring is an alternative to antiviral therapy. In addition, liver biopsy may be repeated in 5 years to assess progression rate (Strader, 2004) in monoinfected patients.

However, the benefits of liver biopsy have to be weighted in the light of sampling error (Regev, 2002) and against the risk of complications such as hemorrhage, puncture of adjoining organs, or mortality. Fatal hemorrhage may occur in up to 0.11% of all liver biopsies, with higher prevalence in malignant liver disease (McGill, 1990; Bravo, 2001).

As the efficacy of HCV therapy has improved, guidelines have been modified so that all patients with HCV should be evaluated for treatment (Strader, 2004; DHHS, 2006). This change in protocol has effectively reduced the need for liver biopsy, but it is still useful in making treatment decisions in select cases, such as patients with difficult to treat genotypes. For patients with genotypes 1 and 4, therapy should be individualized based on severity of liver disease, as determined by either histology or clinical and laboratory signs of cirrhosis, such as spider angiomas, splenomegaly, jaundice, ascites, encephalopathy, hyperbilirubinemia, hypoalbuminemia, thrombocytopenia, and prolonged prothrombin time (PT)/increased international normalized ratio (INR). However, the latter are signs of advanced liver disease, which may be avoided with earlier treatment.

Alternatives to biopsy, such as FibroScan® (Echosens, France) and panels of serum markers of fibrosis (eg, FibroTest, Biopredictive; FibroTest, Biopredictive), may eventually obviate the need for biopsy in all patients. Among these, FibroScan® determines liver stiffness, which is a noninvasive measure of fibrosis (Ziol, 2005), while FibroTest (Imbert-Bismut, 2001) and FibroTest® (Christensen, 2006) are composite scores of serum markers of inflammation and fibrosis. These noninvasive methods are more accurate in predicting very mild fibrosis or cirrhosis and may help avert liver biopsy for patients whose liver histology may be at the extremes of the spectrum of inflammation and fibrosis.

### Management

**ALL PATIENTS WITH HCV SHOULD ALSO BE TESTED FOR HEPATITIS A VIRUS (HAV), HBV, AND HIV. If negative, patients should be vaccinated against both HBV and HAV. The immune response to HAV vaccine is good, even in relatively immune-suppressed individuals, while successful HBV vaccination relies on relatively high CD4 counts (>200 per mL or if CD4+ levels rise above 500 per mm³ in the prior nonresponder (Laurence, 2005).**

Treatment of chronic HCV has considerably improved over the last 10 years. A substantial proportion of patients can achieve serum viral eradication, although current treatments have limitations.

Comorbidities such as diabetes, cardiovascular and kidney disease, psychiatric history, and hematologic abnormalities must be considered before initiating treatment (Strader, 2004).

Pretreatment retinal examination is important, since a rare neuroretinitis (seen more often in patients with diabetes) may occur that is a medical emergency, necessitating discontinuation of treatment.

Alpha interferon is difficult to apply in decompensated cirrhosis (Child-Pugh class C) and may precipitate deterioration. Patients with signs of decompensated cirrhosis, such as jaundice, marked coagulopathy, spontaneous bacterial peritonitis, variceal bleeding and/or hepatic encephalopathy (Table 2) meeting Child-Pugh class C (Table 3) should be considered for liver transplantation.

A majority of coinfected patients, with normal ALT, have mild fibrosis, while 30% may have moderate stage 2 fibrosis (Sanchez-Conde, 2006). Noninvasive markers are less accurate in coinfected patients (Macias, 2006; Cacoub, 2006; Bourliere, 2006). A majority of coinfected patients with normal ALT have mild fibrosis, while up to 30% have moderate stage 2 fibrosis (Sanchez-Conde, 2006). Untreated coinfected patients may need to undergo liver biopsy more frequently than every 5 years. Over a 3-year time interval, a 2-stage progression has been noted on liver biopsy in 22% of patients with stage 1 fibrosis at baseline. Faster progression has been linked to persistently elevated AST and ALT (>100 IU/mL) (Sulkowski, 2006).

### TABLE 1. Assays for Quantification of HCV RNA in Serum

<table>
<thead>
<tr>
<th>Assay</th>
<th>1 IU/L Conversion</th>
<th>Technique</th>
<th>Dynamic Range (IU/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Versant HCV RNA version # 3.0 Quantitative Assay</td>
<td>5.2 copies/mL</td>
<td>Semi-automated branched DNA assay</td>
<td>615–700,000</td>
</tr>
<tr>
<td>LCX HCV RNA Quantitative Assay</td>
<td>3.8 copies/mL</td>
<td>Semi-automated competitive RT-PCR</td>
<td>25–2,630,000</td>
</tr>
<tr>
<td>SuperQuant</td>
<td>3.4 copies/mL</td>
<td>Semi-automated competitive RT-PCR</td>
<td>30–1,470,000</td>
</tr>
</tbody>
</table>

Adapted from Pawlotsky JM. *Gastroenterology.* 2002;122:1554-1568.
TABLE 2. Grading of Hepatic Encephalopathy

<table>
<thead>
<tr>
<th>Grade</th>
<th>Level of Consciousness</th>
<th>Intellectual Function</th>
<th>Neurologic Findings</th>
<th>EEG</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Lack of awareness</td>
<td>Short attention span, forgetfulness, mild confusion, agitation</td>
<td>Uncoordination</td>
<td>Slowing (5–6 cps)</td>
</tr>
<tr>
<td></td>
<td>Personality change</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Day/night sleep pattern reversal</td>
<td></td>
<td>Mild asterixis tremor</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>Lethargic</td>
<td>Disoriented</td>
<td>Asterixis</td>
<td>Slowing</td>
</tr>
<tr>
<td></td>
<td>Inappropriate behavior</td>
<td></td>
<td>Hyperactive reflexes</td>
<td>Tristribic</td>
</tr>
<tr>
<td>3</td>
<td>Asleep</td>
<td>Loss of meaningful communication</td>
<td>Asterixis</td>
<td>Slowing</td>
</tr>
<tr>
<td></td>
<td>Rous edness</td>
<td></td>
<td>Hyperactive reflexes</td>
<td>Tristribic</td>
</tr>
<tr>
<td>4</td>
<td>Comatose</td>
<td>Absent</td>
<td>Decerebrate</td>
<td>Very slow (2–3 cps), delta</td>
</tr>
</tbody>
</table>


TABLE 3. Child-Pugh Score Calculation and Interpretation

<table>
<thead>
<tr>
<th>Child-Pugh Score Interpretation: Points</th>
</tr>
</thead>
<tbody>
<tr>
<td>Class A: 5–6</td>
</tr>
<tr>
<td>Class B: 7–9</td>
</tr>
<tr>
<td>Class C: 10–15</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Points</th>
<th>1</th>
<th>2</th>
<th>3</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bilirubin</td>
<td>&lt;2.0</td>
<td>2.0–3.0</td>
<td>&gt;3.0</td>
</tr>
<tr>
<td>Albumin</td>
<td>&gt;3.5</td>
<td>3.5–2.8</td>
<td>&lt;2.8</td>
</tr>
<tr>
<td>PT prolongation (&lt;1NR):</td>
<td>&lt;4 seconds (&lt;1.7)</td>
<td>4–6 seconds (1.7–2.3)</td>
<td>&gt;6 seconds (&gt;2.3)</td>
</tr>
<tr>
<td>Ascites</td>
<td>Absent</td>
<td>Mild-Moderate</td>
<td>Severe/Refractory</td>
</tr>
<tr>
<td>Encephalopathy</td>
<td>Absent</td>
<td>Mild (1–2)</td>
<td>Severe (3–4)</td>
</tr>
</tbody>
</table>

Points should be ascribed according to clinical and laboratory data and then added to establish Child-Pugh score and interpretation as stated above.

Combination Therapy with IFN alpha and Ribavirin

THE IFNS ARE A SYSTEM OF RELATED PROTEINS DERIVED FROM A MULTIGENE family that work via species-specific surface cell receptors. Interferon monotherapy induces a nonspecific cytokine response, Th1 and Th2, with sustained responders showing increased Th1 reactivity by augmented IFN-γ production at week 4 (Cramp, 2000).

Pegylated Interferons

LIMITATIONS IN THE EFFECTIVENESS OF IFN-α HAVE BEEN ATTRIBUTED to its rapid systemic clearance and short plasma elimination half-life (t½) of about 8 hours. Pegylated (PEG) subcutaneous formulations of IFN-α have been developed by covalent attachment of recombinant IFN-α to branched 40- or 12-kD polyethylene glycol moieties (Kozlowski, 2001). These molecules protect the IFN protein from enzymatic degradation, thus reducing systemic clearance. Pegylation alters the pharmacokinetics and pharmacodynamics of IFN-α, leading to improved drug concentrations and sustained exposure. Pegylated IFNs have decreased volume of distribution (Vd) and a greater reduction in renal clearance compared with standard IFN.

Two versions, a 12-kD PEG IFN-α 2b (Peg-Intron®, Virafon PEG®, Schering) and a 40-kD PEG IFN-α 2a (Pegasys®, Roche) are approved for the treatment of hepatitis C, but only PEG IFN-α 2a is currently approved by the FDA for the treatment of HCV in HIV-coinfected patients.

Although the 2 available PEG IFN products have different pharmacokinetic profiles and molecular structures, there do not appear to be major differences in their efficacy in the treatment of HCV monoinfection and HCV-HIV coinfection. However, no head-to-head studies have been conducted.

The aim of therapy is to achieve a sustained virological response (SVR: an undetectable RNA 6 months after completion of therapy) and ultimately to reduce hepatic inflammation and severity of fibrosis. Response to treatment depends on genotype, baseline HCV RNA, and dosing and duration of treatment, age, and the stage of fibrosis, as well as clinical factors, such as comorbid conditions. For monoinfected patients, 48 weeks of PEG IFN-α 2b (1.5 µg/kg/wk) in combination with weight-based ribavirin (800 mg/d) achieved an overall SVR of 54% (42% for genotype 1 and 80% for genotypes 2 and 3), in comparison to an overall SVR of 47% with standard IFN-α 2b plus ribavirin. Analysis of response by weight-based dosing demonstrated that the optimum dose of ribavirin is 10.6 mg per kg per day, and those receiving this dose achieved an SVR of 48% for genotype 1 and 88% for genotypes 2 and 3 (Manns, 2001; Hadziyannis, 2004). Similar results have been obtained with PEG IFN-α 2a plus a fixed dose of ribavirin (Fried, 2002). For those monoinfected with genotypes 2 or 3, a reduced dose (800 mg/d) of ribavirin was adequate, as was only 24 weeks of combination therapy with PEG IFN and ribavirin (Hadziyannis, 2004). This finding has been extrapolated and applied to the use of PEG IFN-α 2b and ribavirin.

Ribavirin

RIBAVIRIN, A GUanosine nucleoside analogue, shows only modest activity against HCV but increases the activity of standard and PEG IFN-α when the 2 are used in combination. The precise mode of action probably includes perturbation of intracellular nucleoside triphosphate pools. Ribavirin may also induce mutations in the HCV genome, affecting viral replication (Dixit, 2004).
**Current Treatment Protocols**

**Monoinfected Patients**

The aim of therapy is to achieve an undetectable HCV RNA 6 months following therapy (SVR) and ultimately to reduce hepatic inflammation and severity of fibrosis. Pegylated IFN-α 2b is administered at a dose of 1.5 μg per kg per week by subcutaneous injection. With this form of PEG IFN, ribavirin is coadministered according to the body weight of the patient for all genotypes in divided, oral daily doses. For patients who weigh less than 65 kg, the dose of ribavirin is 800 mg; for patients 65 to 85 kg, the dose is 1000 mg; and for patients over 85 kg, the dose is 1200 mg per day, divided in 2 doses. Pegylated IFN-α 2a (Pegasys®) is given at a fixed dose of 180 μg per week. Coadministration of ribavirin in patients with genotype 1 is 1000 mg for those less than 75 kg and 1200 mg for those equal to or greater than 75 kg, in divided daily doses. Patients with genotypes 2 and 3 are treated with 800 mg of ribavirin. For genotypes 4, 5, and 6, there is insufficient evidence for treatment regimens and therapy needs to be individualized.

The pivotal studies suggested that a 24-week schedule for genotype 2 or 3 HCV-monoinfected patients is sufficient, whereas patients with genotype 1 require 48 weeks of therapy. Few patients with genotype 4 have been studied, but an SVR of 34% has been reported after a 48-week course of combination therapy, indicating that it should be treated in a similar manner to genotype 1 (Derbala, 2005). There are limited published data on treatment outcomes in patients with genotypes 5 and 6.

Patients with genotype 1 who do not show an early viral response (EVR; HCV RNA decline of at least 2 log₁₀ or undetectable HCV RNA after 12 weeks of therapy determined by quantitative PCR) have little opportunity of achieving an SVR, and therapy should be discontinued. For patients who achieve an EVR, HCV RNA should be measured at week 24. Patients who have detectable HCV RNA positive should stop therapy. Patients with genotype 2 or 3 who have a high response rate to treatment (Strader, 2004) and, in those patients, HCV RNA testing at week 12 may not be cost-effective. However, testing at week 12 may avoid unnecessary medication for nonresponders and may help to motivate patients who are responding to stay on treatment.

The most common, early adverse reactions of IFN include an influenza-like syndrome: chills, fever, malaise, muscle aches, and headaches. These symptoms may be ameliorated by acetaminophen (paracetamol), water, and exercise. Poor appetite, weight loss, increased somnolence, psychologic effects (irritability, anxiety, depression), hair loss, thomboctopenia, and leukopenia are also common adverse effects.

Mild depression is not unusual and can often be treated with selective serotonin reuptake inhibitors (SSRIs), but a careful, pretreatment psychologic inventory is necessary to determine those who are at risk for developing severe depression. These patients may need structured psychologic support, which may delay or even preclude HCV treatment.

Dose reductions of IFN and/or ribavirin may be necessary, particularly in cirrhotic patients with low white cell and platelet counts due to portal hypertension and splenic sequestration. Unusual and severe adverse effects include seizures, acute psychoses, bacterial infections, and autoimmune reactions. Thyroid disease (both hyperthyroidism and hypothyroidism) is relatively common, can be seen in up to 5% of patients taking IFN, and can be permanent, requiring long-term therapy. Thyroid disease usually occurs in the setting of pre-existing antithyroid antibodies. Proteinuria, cardiomyopathy, skin rashes, interstitial lung disease, bone marrow suppression/aplasia and antibodies against IFN may also develop.

Although adverse effects are common, most are reversible and will resolve upon discontinuation of therapy. Occasionally, specialty referral is indicated for unusual reactions and growth factor supplementation can also be instituted to avoid dose reduction or discontinuation due to anemia or leukopenia.

The major adverse effects of ribavirin are dose-dependent and include: reversible hemolytic anemia, myalgia (muscle pain), hypouricemia, dyspepsia, and irritability. Patients should be carefully monitored for the above adverse effects clinically and serologically with CBCS, AST/ALT, uric acid, albumin, bilirubin, and thyroid function tests measured at least every 4 weeks. Any abnormal result should prompt immediate consideration and further investigation.

Due to the potential for ribavirin-induced teratogenicity, 2 methods of contraception should be reinforced prior to initiation of therapy and at each treatment-monitoring visit.

Dose reductions of IFN and/or ribavirin may be necessary, particularly in cirrhotic patients who develop anemia and/or leukopenia due to bone marrow suppression, or thrombocytopenia from portal hypertension and splenic sequestration.

Adherence to therapy is an important factor for improving SVR, but adverse effects necessitate dose reductions or discontinuation of therapy in 14% of patients. Aggressive management of adverse effects will help patients remain on HCV therapy.

Supportive therapies such as antidepressants, erythropoietin, and G-CSF have been demonstrated to reduce the incidence of IFN-induced depression, anemia and neutropenia, respectively. In anemic HCV-infected patients treated with ribavirin/IFN, epoetin alfa increases hemoglobin levels and maintains ribavirin dosing (Dieterich, 2003). Although these interventions can improve quality of life and enhance adherence, they have not yet been shown to benefit the SVR rate. Furthermore, they considerably add to the expense of therapy.

In addition to encouragement and support, patients should be advised to minimize or eliminate their intake of alcohol, since studies of interferon monotherapy reported a decrement in response to HCV treatment when alcohol was used, and alcohol consumption has been associated with higher HCV RNA.

Nonresponders to PEG IFN and ribavirin combination therapy, defined as patients whose HCV RNA levels remain stable on treatment, have a poor response to retreatment and, at present, no consensus exists regarding the management of this population. Those who relapse after standard IFN monotherapy, or standard IFN and ribavirin, may achieve an SVR when retreated with PEG IFNs and standard doses of ribavirin, particularly if they are nongenotype 1. As in naïve patients, treatment should be stopped if there is no retreatment EVR. Several major trials, including EPIC3 and HALT-C are currently investigating the long-term use of low-dose PEG IFN in nonresponders and those who relapse (ie, HCV RNA becomes undetectable on treatment but is detected again after discontinuation of therapy).

Current studies are investigating abbreviated courses of treatment for patients with genotype 1 and low viral loads (<400,000 IU/mL) who show a rapid viral response (EVR; negative HCV RNA by PCR at 1 month). Similarly, it may be possible to stop treatment at 16 weeks for patients with genotype 2 and 3 who have an EVR, while longer treatment may be needed for genotype 3 patients with HCV RNA above 800,000 IU/mL (von Wagner, 2005). Optimal dosing of ribavirin may be the key to achieving an EVR, since patients with HCV RNA levels higher than 800,000 IU per mL in genotype 3 may be more likely to relapse. Attractive as they may be, shorter courses of therapy are not universally accepted, since more studies are needed to determine the appropriate duration of treatment. Long-term follow-up on patients with SVR reveals ALT normalization and fibrosis and cirrhosis regression in a majority of patients with persistent HCV RNA in monocytes or liver tissue in extremely isolated cases (Maylin, 2006).
**Coinfected Patients**

Several guidelines have addressed the management of HCV/HIV coinfection (Strader, 2004; DHHS, 2006). Figure 1 provides a practical management algorithm for HIV-HCV coinfected patients.

Treatment is generally more complex in coinfectected patients as a result of myelosuppression, drug interactions, antiretroviral-related hepatotoxicity, and advanced HIV disease. It is accepted that HCV treatment does not influence HIV progression or immune status, as the incidence of AIDS-defining events remained uniformly low, despite transient decrease in CD4 counts, while CD4+ percentage slightly increased (Torriani, 2004). In addition to HCV treatment, optimal immune status and effective HIV control should be achieved in an effort to slow fibrosis progression.

The indications for treatment are generally based on a combination of virologic findings and a full hepatic assessment. Most guidelines indicate that all HIV-positive patients with chronic HCV should be considered candidates for treatment (Strader, 2004; DHHS, 2006). Treatment is strongly recommended for patients with elevated serum aminotransferases, CD4 counts of greater than 350 cells per µl, HIV RNA less than 1000 copies per mL, and no alcohol intake, although the degree of fibrosis needs to be taken into account.

Similar to monoinfected patients, SVR is a strong predictor of SVR as long as dose reductions or discontinuation do not compromise the treatment course, and EVR is required to warrant a full course of therapy.

In a review of treatment practices, 70% of all coinfectected patients were ineligible for HCV treatment, primarily due to substance abuse, nonadherence with medical follow-up, advanced HIV disease, decompensated liver status, or medical comorbidities (Fleming, 2003). However, multidisciplinary models that incorporate mental health care and drug treatment (including methadone and buprenorphine) have successfully delivered HCV treatment to monoinfected and coinfectected patients with multiple diagnoses (Backmund, 2001; Schaefer, 2003; Van Thiel, 2003; Cournot, 2005; Litwin, 2005; Taylor, 2005; Sylvestre, 2005).

If possible, it is recommended that HIV management be optimized before commencing HCV treatment. Interferon may induce a slight decline in absolute CD4 count, but this does not appear to be of clinical significance.

Antiretroviral agents for HIV/HCV coinfectected patients should be carefully selected, since some are more likely to induce hepatotoxicity, depletion of hepatocellular mitochondrial DNA, and hepatic steatosis (Walker, 2004; Sulkowski, 2005; Mc Govern, 2006). Liver enzyme levels should be routinely monitored (Figure 2).

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**Box 1**: Weight-based dosage of ribavirin used with Peg IFN-α 2b

<table>
<thead>
<tr>
<th>Weight, kg</th>
<th>Redipen® µg/0.5 mL</th>
<th>Volume, mL</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt;40</td>
<td>50</td>
<td>0.5</td>
</tr>
<tr>
<td>41–50</td>
<td>80</td>
<td>0.4</td>
</tr>
<tr>
<td>51–64</td>
<td>80</td>
<td>0.5</td>
</tr>
<tr>
<td>65–75</td>
<td>120</td>
<td>0.4</td>
</tr>
<tr>
<td>76–86</td>
<td>120</td>
<td>0.5</td>
</tr>
<tr>
<td>&gt;85</td>
<td>150</td>
<td>0.5</td>
</tr>
</tbody>
</table>

**Box 2**: Weight-based dosage of Peg IFN-α 2b

<table>
<thead>
<tr>
<th>Weight, kg</th>
<th>Peg IFN-α 2b µg/kg</th>
<th>Volume, mL</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt;40</td>
<td>1.5</td>
<td>0.5</td>
</tr>
<tr>
<td>40–64</td>
<td>1.5</td>
<td>0.5</td>
</tr>
<tr>
<td>65–85</td>
<td>1.5</td>
<td>0.5</td>
</tr>
<tr>
<td>86–105</td>
<td>1.5</td>
<td>0.5</td>
</tr>
<tr>
<td>&gt;105</td>
<td>1.5</td>
<td>0.5</td>
</tr>
</tbody>
</table>

**Box 3**: Weight-based dosage of ribavirin used with Peg IFN-α 2a

<table>
<thead>
<tr>
<th>Weight, kg</th>
<th>Peg IFN-α 2a µg/kg</th>
<th>Volume, mL</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt;75</td>
<td>80</td>
<td>0.4</td>
</tr>
<tr>
<td>&gt;75</td>
<td>120</td>
<td>0.5</td>
</tr>
</tbody>
</table>

*Trials to optimize response rates in mono- and coinfectected patients are ongoing. Results from the PRESCO trial indicate that weight-based ribavirin dosing, regardless of genotype, is associated with SVR in coinfectected patients.
Coadministration of didanosine (ddI) and ribavirin should be avoided because of an increased risk of lactic acidosis (Lafeuillade, 2001) and/or pancreatitis (Salmon-Ceron, 2001), while aztidin conjunction with ribavirin increases the risk of anemia (Alvarez, 2006).

Use of stavudine (d4T) during hcv treatment is associated with weight loss and lipoatrophy (Perez Olmeda, 2003). As with hcv monoinfection, the primary goal of hcv treatment for coinfected patients is svr. However, even in the absence of svr, benefits of treatment have been shown and may include: reduction in the risk of hcc (Soriano, 2004); reduction in the risk of haart-associated hepatotoxicity (Uberti-Foppa, 2003); and delay or reversal of fibrosis progression (Lissen, 2006).

The treatment regimen is the same as in monoinfected patients: peg ifn and ribavirin. Although svr is less likely for coinfected patients, if hiv is well controlled and hcv therapy is tailored by response to treatment and weight-based ribavirin dosing, response can be optimized. Four randomized, controlled trials have been published that examine the efficacy of peg ifn and ribavirin for treatment of hcv in patients coinfected with hiv (Tables 4 and 5). In general, the mean or median cd4 counts were high and more than 80% were undergoing haart. Response rates ranged from 27% to 40%, which was better than that seen with standard ifn (12%–21%), but worse than that observed in monoinfected patients undergoing combination therapy (Carrat, 2004; Chung, 2004; Laguno, 2004; Torriani, 2004).

In coinfected genotype and baseline rna also exerted a strong effect upon response rates. Higher svrs were observed in patients with genotypes 2 and 3, as well as in patients with genotype 1 and lower viral loads. Better outcomes were also noted in young patients (aged younger than 40 years) without cirrhosis, those with elevated aminotransferases, and those with low or undetectable hiv rna. In the apricot study, a

| HCV, hepatitis C virus; PCR, polymerase chain reaction; SSRI, selective serotonin reuptake inhibitor |

Higher doses (up to 1200 mg) are approved for monoinfected patients, and some coinfected patients may derive similar benefit from a more aggressive regimen. Furthermore, APRICOT found that coinfected patients have a higher rate of relapse and may need 48 weeks of treatment, regardless of HCV genotype. As with HCV monoinfection, coinfected patients who fail to achieve an SVR have little, if any chance of achieving an SVR (Ballesteros, 2004; Laguno, 2007).

HCV treatment may have histologic benefits, even in the absence of SVR. Studies based on paired liver biopsies, taken before and after HCV treatment, showed improved histology in a majority of treatment responders and in up to 40% of patients who did not achieve SVR (Lissen, 2006; Soriano, 2006).

In HIV-coinfected patients, SVR appears to be equally durable. Soriano and colleagues reported no HCV rebound among 77 patients who achieved SVR (minimum 6 month and mean of 58 months of long term follow-up). In addition, no liver decompensation or HCC are in this analysis (Soriano, 2004).

Liver transplantation in HCV-suppressed coinfected patients has become an option for decompensated liver disease (see http://clinicaltrials.gov/ct/show/NCT00074386?order=1). However, survival beyond 4 years was noted to be poorer than in HCV monoinfected, and related to pretransplant Model for End-stage Liver Disease (MELD) score and to posttransplant HAART intolerance, presence of HIV RNA above 400 and CD4 counts below 200 (Ragni, 2003; Duclos, 2006). In addition, HCV had a more severe course after orthotopic liver transplantation (OLT) (Duclos, 2006). It is likely that patient selection for current treatments will be based on prediction models, in establishing risk-benefit ratio, in difficult-to-treat cases, such as genotype 1 coinfected patients, with advanced fibrosis.

### TABLE 4. PEG IFN + Ribavirin Trials in HIV-HCV Coinfected Patients

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Chung, 2004 ACTG</th>
<th>Torriani, 2004 APRICOT</th>
<th>Carrat, 2004 RIBAVIC</th>
<th>Laguno, 2004</th>
</tr>
</thead>
<tbody>
<tr>
<td>Location</td>
<td>United States</td>
<td>International</td>
<td>France</td>
<td>Spain</td>
</tr>
<tr>
<td>Design</td>
<td>PEG 2a + RBV VS</td>
<td>PEG 2a + RBV VS PEG 2a + placebo VS STD 2a + RBV</td>
<td>PEG 2b + RBV VS STD 2b + RBV</td>
<td>PEG 2b + RBV VS STD 2b + RBV</td>
</tr>
<tr>
<td>PEG Dose</td>
<td>180 µg</td>
<td>180 µg</td>
<td>1.5 µg/kg</td>
<td>100–150 µg</td>
</tr>
<tr>
<td>Ribavirin</td>
<td>600 mg/d starting dose in 200-mg monthly increments up to a total dose of 1000 mg/d</td>
<td>800 mg/d</td>
<td>800 mg/d</td>
<td>800–1200 mg/d</td>
</tr>
<tr>
<td>Weeks</td>
<td>48</td>
<td>48</td>
<td>48</td>
<td>48</td>
</tr>
<tr>
<td>Undetectable HIV RNA</td>
<td>61%</td>
<td>60%</td>
<td>70% &lt;400</td>
<td>70% &lt;200</td>
</tr>
<tr>
<td>HCV RNA &gt; 800,00 IU</td>
<td>83%</td>
<td>72%</td>
<td>Not reported</td>
<td>47%</td>
</tr>
<tr>
<td>Genotype 1</td>
<td>77%</td>
<td>61%</td>
<td>48%</td>
<td>55%</td>
</tr>
<tr>
<td>Fibrosis Cirrhosis</td>
<td>11%</td>
<td>12%</td>
<td>39%</td>
<td>29%</td>
</tr>
<tr>
<td>Adverse Effects Management</td>
<td>PEG/RBV dose reduction</td>
<td>PEG/RBV dose reduction Erythropoietin G-CSF</td>
<td>PEG/RBV dose reduction</td>
<td>PEG/RBV dose reduction</td>
</tr>
</tbody>
</table>

**RBV, ribavirin; PEG, pegylated interferon; G-CSF, granulocyte-colony stimulating factor; STD, standard interferon.**

### TABLE 5. Response in Coinfected Patients

<table>
<thead>
<tr>
<th>Reference</th>
<th>Genotype 1 SVR</th>
<th>Genotype 2 and 3 SVR</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chung, 2004 ACTG</td>
<td>PEG 2a + RBV 14%</td>
<td>73%</td>
</tr>
<tr>
<td></td>
<td>STD 2a + RBV 6%</td>
<td>33%</td>
</tr>
<tr>
<td>Torriani, 2004 APRICOT</td>
<td>PEG 2a + RBV 29%</td>
<td>62%</td>
</tr>
<tr>
<td></td>
<td>STD 2a + RBV 7%</td>
<td>20%</td>
</tr>
<tr>
<td>Carrat, 2004 RIBAVIC</td>
<td>PEG 2b + RBV 17%</td>
<td>44%</td>
</tr>
<tr>
<td></td>
<td>STD 2b + RBV 6%</td>
<td>43%</td>
</tr>
<tr>
<td>Laguno, 2004</td>
<td>PEG 2b + RBV WB 38%</td>
<td>53%</td>
</tr>
<tr>
<td></td>
<td>STD 2b + RBV 7%</td>
<td>47%</td>
</tr>
</tbody>
</table>

**PEG, pegylated interferon; RBV, ribavirin; WB, weight based.**

Baseline CD4 percentage higher than 19.1%, and baseline HCV RNA below 400,000 IU per mL predicted higher SVR in genotype I patients (Dieterich, 2006). In addition, an early increase in CD4 cell percentage, after 1 month of treatment, was found to predict viral response (Neau-Cransac, 2005).

Differences in SVR rates among the studies include patient characteristics—such as general health status and race—and study design and ribavirin dose. It is possible that higher relapse rates occurred due to a lower dose of ribavirin (800 mg).

Questions remain regarding best dosing and duration of HCV treatment in HIV/HCV coinfected patients. In February of 2006, PEG IFN-α 2a with 800 mg of ribavirin was approved for HIV/HCV coinfected patients based on data from the APRICOT Study Group Trial (Torriani, 2004). The study conservatively dosed ribavirin at 800 mg daily in an attempt to prevent dose reduction or discontinuation due to anemia. Discontinuation of treatment was observed in 12% to 17% of patients and supplemental hemopoietic factors were required in 60% of patients taking zidovudine who developed anemia.
New Strategies and Treatments on the Horizon

IN VITRO, HCV IS EXCEEDINGLY DIFFICULT TO CULTURE, BUT THE RECENT development of an HCV replicon system, and the production of robust cell culture models allow for a more detailed investigation of HCV replication (Lindenberg, 2005; Wakita, 2005). Currently, there is a robust pipeline in clinical and preclinical development.

In HCV monoinfection, different treatment strategies are being explored. Extension of treatment with peg IFN-α 2a plus ribavirin from 48 to 72 weeks considerably increases the rate of SVR in patients with detectable viremia at week 4 of treatment (Sanchez-Tapias, 2006). Treatment of overweight and obese patients could achieve SVR rates similar to lean subjects by dosing ribavirin using a weight-based scheme (1200 mg/day for weight 85–105 kg, 1400 mg/day for 105–125 kg) (Jacobson, 2006). Concerns about inducing anemia with higher ribavirin doses were addressed in a recent study, showing that pre-emptive treatment with erythropoietin at the onset of combination therapy with peg IFN-α 2b and high-dose ribavirin (>13 mg/kg), significantly increased SVR and decreased the incidence of anemia (Shiffman, 2005). It also allowed maintenance of higher doses of ribavirin, while preserving quality of life (Sulkowski, 2005).

In HIV/HCV coinfection, higher doses of ribavirin (1000 mg/day for patients weighing less than 75 kg and 1200 mg/day for those more than 75 kg) in addition to peg IFN-α 2a, for 48 weeks, led to better SVR rates for all genotypes, when compared with historic APRICOT data. However, prolonging treatment to 72 weeks for genotype 1 led to more adverse effects and to treatment discontinuation (Nunez, 2006), likely negating the slight increase in SVR.

Long-term, low-dose peg IFN-α 2b led to improvements in fibrosis and inflammation scores in HCV nonresponders with advanced fibrosis or cirrhosis (Kaiser, 2006).

Two trials, Slam C and HRN 004, are exploring peg IFN maintenance in coinfected patients with advanced fibrosis who failed current treatment regimens, and are addressed towards stopping fibrosis progression.

Important progress is being made in the development of new treatments, particularly in new specific inhibitors of hepatitis C. Infergen® (Valent Pharmaceuticals), also known as consensus IFN (IFN alphacon-1), appears to be a promising therapy for monoinfected and coinfected nonresponders (Bacon, 2006; Leevy, 2005). In a difficult-to-treat group with genotype 1 and advanced fibrosis, consensus interferon led to an end-of-treatment response of 19% with 15 µg per day and ribavirin (Bacon, 2006).

Viramidine® (taribavirin hydrochloride; Valent Pharmaceuticals), a prodrug of ribavirin, may cause less hemolytic anemia due to its preferential uptake by the liver, which effectively reduces the plasma concentration of ribavirin and exposure to red blood cells. Phase 3 studies using 600 mg twice daily have shown that, although taribavirin results in a lower incidence of anemia, it is less effective than weight-based ribavirin. Recently, it was shown that achieving taribavirin levels higher than 18 mg per kg yields an SVR rate similar to that of standard ribavirin treatment, with a lower rate of anemia (Jacobson, 2006). A phase 2 trial will be looking at safety and efficacy of weight-based taribavirin.

Albuferon® (albinterferon alpha 2b; Human Genome Sciences), which is currently in phase 2 and 3 trials, is an 85.7-kD protein consisting of IFN-α that is genetically fused to human serum albumin. This process extends the half-life of the medication, allowing for dosing intervals of up to 2 to 4 weeks. In a recent report, nonresponders to prior IFN-α underwent treatment with a combination of albinterferon alpha 2b and weight-based ribavirin. This led to an SVR rate of 28%, at a dose of 900 mg bi-weekly, with 15% of genotype 1 patients experiencing an SVR at 1200 mg monthly (Nelson, 2006).

Several new enzymatic inhibitors of hepatitis C virus show some promise in phase 1 and 2 studies. Valopicitobine (Idenix Pharmaceuticals), or NM281, is a prodrug of the polymerase inhibitor MN107, which competitively inhibits HCV RNA polymerase. As an orally administered daily dose, valopicitobine causes a rapid decline in HCV RNA concentrations in animal models, with a mean viral load reduction after 7 days of 1.05 log_{10} and 0.83 log_{10} in the high- and low-dose groups, respectively. Results from a phase 2 study of IFN and valopicitobine in nonresponders were disappointing, since none achieved SVR; final results from a trial in naïve patients are expected in mid-2007 (Afdal, 2007).

The protease enzymes may represent dual therapeutic agents as they suppress virus replication and improve host IFN responsiveness. VX-950 (Vertex Pharmaceuticals), a peptidomimetic protease inhibitor of the hepatitis C NS3/4A protease, has shown promise in phase 1 and 2 studies, but genomic analysis of HCV sequences after only a few weeks of monotherapy have shown a rapid emergence of resistance. A triple-drug, phase 2 study of VX-950 (Vertex Pharmaceuticals) (in combination with peg IFN and ribavirin) resulted in undetectable HCV RNA (<10 IU/mL) in all 12 patients after 28 days of treatment (Lawitz, 2006). Further studies are ongoing to assess the efficacy of 12- and 24-week treatment of VX950 at a dose of 750 mg every 8 hours. The study designs include combination protocols of the protease inhibitor with either peg IFN alone (double therapy) or peg IFN and ribavirin (triple therapy). The ketoamide SCH 503034 (Schering-Plough Corporation) is another peptidomimetic protease inhibitor that was shown to inhibit HCV replication in vitro. Phase 2 studies in combination with IFN-α 2b and, now, ribavirin, are in progress. Future use of HCV protease inhibitors in PI-exposed HIV-positive patients, may be impacted by the emerging mutations of NS3 protease domain, leading to HCV-PI resistance (Morsica, 2006).

Other novel drugs that are being tested include nucleoside-resistant ribozymes, which inhibit RNA translation, and antisense inhibitors, which reduce production of proteins necessary for HCV replication. An inhibitor of hepatocyte apoptosis—PF-0399—was reported to markedly decrease ALT and AST in HCV patients with advanced fibrosis. Histology outcomes need to be studied (Shiffman, 2006). A cyclophyllin antagonist produced an almost 4-log drop in HCV RNA and 1-log drop in HIV RNA, after 15 days, in a phase 1 trial in coinfect patients (Flisiak, 2006).

Similarly, several vaccines were tested, designed to stimulate an immune response to envelope proteins, with antiviral and antifibrotic effects (Leroux-Roels, 2004; DiBisceglie, 2005). As an adjunctive therapy, interferon will continue to be the backbone of treatment for some time. In the future, HAART-like triple-therapy may prove to increase efficacy and tolerability, and, hopefully, shorten the duration of treatment. It is likely that resistance testing will guide treatment decisions using new antiviral agents. Trials of new treatment strategies involving current drugs and new agents in coinfect patients should expand the understanding of the most appropriate treatment protocols in this population.


